



# HSV1/2 Genital Infection in Mice Cause Reversible Delayed Gastrointestinal Transit: A Model for Enteric Myopathy

Arun Chaudhury<sup>1\*</sup>, Vijaya Sena Reddy Dendi<sup>2</sup>, Mousumi Chaudhury<sup>1</sup>, Astha Jain<sup>3</sup>, Madhukar Reddy Kasarla<sup>4</sup>, Kiran Panuganti<sup>5</sup>, Gaurav Jain<sup>6</sup>, Abhijit Ramanujam<sup>7</sup>, Bhavin Rena<sup>8</sup>, Sudheer Reddy Koyagura<sup>9</sup>, Sumit Fogla<sup>10</sup>, Sunil Kumar<sup>11</sup>, Nawal Singh Shekhawat<sup>12</sup> and Srinivas Maddur<sup>13,14</sup>

## **OPEN ACCESS**

## Edited by:

Achim Weber, Universität Zürich, Switzerland

#### Reviewed by:

Maximilian Ackermann, Universitätsmedizin Mainz, Germany Pengyue Zhang, Indiana University Bloomington, United States Tatsuo Suzutani, Fukushima Medical University, Japan Roopali Mittal, University of Oklahoma Health Sciences Center, United States

\*Correspondence: Arun Chaudhury arunchaudhury.boston@gmail.com

#### Specialty section:

This article was submitted to Pathology, a section of the journal Frontiers in Medicine

Received: 20 January 2017 Accepted: 21 May 2018 Published: 17 July 2018

### Citation:

Chaudhury A, Dendi VSR, Chaudhury M, Jain A, Kasarla MR, Panuganti K, Jain G, Ramanujam A, Rena B, Koyagura SR, Fogla S, Kumar S, Shekhawat NS and Maddur S (2018) HSV1/2 Genital Infection in Mice Cause Reversible Delayed Gastrointestinal Transit: A Model for Enteric Myopathy. Front. Med. 5:176. doi: 10.3389/fmed.2018.00176 <sup>1</sup> GIM Foundation, Little Rock, AR, United States, <sup>2</sup> Christus Trinity Mother Frances Hospital, Tyler, TX, United States, <sup>3</sup> Wanderful Media/University of Southern California, Los Angeles, CA, United States, <sup>4</sup> Parkway Surgical and Cardiovascular Hospital, Fort Worth, TX, United States, <sup>5</sup> Presbyterian Hospital, Denton, TX, United States, <sup>6</sup> Berkshire Medical Center, Pittsfield, MA, United States, <sup>7</sup> Woodland Memorial Hospital, Sacramento, CA, United States, <sup>8</sup> Xenco Laboratories, Houston, TX, United States, <sup>9</sup> Northwest Medical Center, Bentonville, AR, United States, <sup>10</sup> Beaumont Hospital, Grosse Pointe, MI, United States, <sup>11</sup> Neshoba County General Hospital, Philadelphia, MS, United States, <sup>12</sup> Baptist Hospital, Conway, AR, United States, <sup>13</sup> All India Institute of Medical Sciences, New Delhi, India, <sup>14</sup> ESIC Medical College, Sanathnagar, India

In an interesting investigation by Khoury-Hanold et al. (1), genital infection of mice with herpes simplex virus 1 (HSV1) were reported to cause multiple pelvic organ involvement and obstruction. A small subset of mice succumbed after the first week of HSV1 infection. The authors inferred that the mice died due to toxic megacolon. In a severe form of mechanical and/or functional obstruction involving gross dilation of the colon and profound toxemia, the presentation is called "toxic megacolon." The representative observations by Khoury-Hanold likely do not resemble toxic megacolon. The colon was only slightly dilated and benign appearing. Importantly, HSV1 infection affected the postjunctional mechanisms of smooth muscle relaxation like the sildenafil-response proteins, which may have been responsible for defective nitrergic neurotransmission and the delayed transit. Orally administered polyethylene glycol reversed the gastrointestinal "obstruction," suggesting a mild functional type of slowed luminal transit, resembling constipation, rather than toxic megacolon, which cannot be reversed by an osmotic laxative without perforating the gut. The authors suggest that the mice did not develop HSV1 encephalitis, the commonly known cause of mortality. The premature death of some of the mice could be related to the bladder outlet obstruction, whose backflow effects may alter renal function, electrolyte abnormalities and death. Muscle strip recordings of mechanical relaxation after electrical field stimulation of gastrointestinal, urinary bladder or cavernosal tissues shall help obtain objective quantitative evidence of whether HSV infection indeed cause pelvic multi-organ dysfunction and impairment of autonomic neurotransmission and postjunctional electromechanical relaxation mechanisms of these organs.

Keywords: enteric nervous system, nitrergic neurotransmission, virus, guanylyl cyclase, megacolon

1

In a recent study, Khoury-Hanold et al. (1) investigates a topic of significant current epidemiological significance viz., genital infection of HSV1, usually an oro-labiotropic virus in mice (1). The authors demonstrate that HSV1 genital infection cause multiorgan autonomic dysfunction, involving the pelvic organs including the urinary bladder and the terminal intestinal tract. The findings are of significant interest, as the study suggests that HSV1 likely affects post-junctional mechanisms of nitrergic downstream signaling that mediate intestinal smooth muscle relaxation.

The authors suggest that transvaginal deliver of HSV1 causes toxic megacolon (1). However, the gastrointestinal phenotype that developed a few days after the infection does not resemble the presentations of "toxic megacolon." Toxic megacolon is a rare but fatal condition that develops unpredictably and rapidly in certain cases of fulminant colitis. This colitis may result from inflammatory or infectious causes. Rarely, toxic megacolon may develop in hereditary conditions, for example as demonstrated in piebald mice, a model of Hirschsprung disease (2). Megacecum and megacolon often develop slowly in animals including models for Hirschsprung disease (3, 4). Pre-existing inflammation may alter luminal environment and facilitate viral entry, for example like cytomegalovirus in ulcerative colitis (5). Toxic megacolon may develop in colitis developing in concomitant Clostridium difficile infection or in subjects with ulcerative colitis (6). This has been examined in animal models of infection and inflammation involving mice, Syrian hamsters and piglets (7). Importantly, systemic toxicity and clinical signs of sepsis are observed, along with massive dilation of colon. In order for reliable pathognomonic diagnosis of toxic megacolon in mice, it is pertinent that two levels of evidence should be presented. First, dilation of small intestinal or colonic dilation must be demonstrated unambiguously, as has been shown in previous studies (Figures 1, 2) (8). Second, evidence of toxicity needs to be presented (Figures 1, 2) (7). Megacolon developing slowly may still show massive dilation of intestines as in Chagas' disease, but does not show local or systemic signs of toxicity (Figure 3) (19). Local signs of toxicity importantly involve ischemia, necrosis or hemorrhage in the wall of the intestine, resulting in unhealthy, angry looking intestinal or colonic loops and obviously, dilation (Figure 1) (7). It may be noted that normal intestinal loops may contain semisolid slurry, not always discrete fecal pellets, or chyme in transit, which may slightly dilate the intestines (Figure 1). This may not be interpreted as megacolon.

Khoury-Hanold et al. (1) present incomplete evidence, which does not provide unequivocal demonstration of either megacolon or sepsis. Only a segment of the large intestine with the terminal anus has been shown (1). The upper part of the segment remains filled with colonic slurry. It is possible that the unfilled segment shows the potentially ganglion-depleted segment, which might have arose from the invasion of the myenteric ganglia by HSV1 in that segment. However, the segment containing the colonic slurry is only slightly gorged; this happens normally when chyle from the cecum enters the colon. This type of mild distention is also seen when chyme/chyle transitions through the small intestines during normal intestinal transit. Lack of demonstration of the entire segment of the colon along with the cecum (1) makes the interpretation of megacolon difficult and unconvincing. It is not mentioned whether the colon was dissected open and cleared of feces prior to estimating its weight and normalizing to total body mass. How the study identified the pre-moribund mice is also not clear while estimating the colonic mass. Some of the mice infected with the neuroattenuated strain  $\Delta 68$ H showed decreased survival. In earlier studies of mice models of megacolon, there has been clear demonstration of the aganglionic colon, the transition zone and the proximal segment, which by overwork, massively dilates (**Figure 2**) (9, 12–14).

In toxic megacolon, the segment of the colon proximal to the obstruction remains massively dilated (mostly with gas, but also with feces), which can be detected easily through radiologic imaging like X-ray abdomen or CT scan (15, 22, 23) (Figure 3). In humans, toxic megacolon involves dilation greater than 6 cm of colon (24). The authors arbitrarily assume a two-fold increase in mass of the colon as their marker for megacolon (1), but it should be remembered that the colon has the intrinsic capacity to dilate to accommodate the contents from the cecum (Figure 1). Demonstration of colonic compliance by demonstrating high volumes at low filling pressures or low colonic tone would be more objective ways to demonstrate true megacolon (25). What is rather important observation is the colonic slurry, rather than formed fecal pellets (1). This may potentially indicate a reduction of colonic mucosal absorptive function, which is possible due to the viral mucositis. Alternatively, this observation could also be normal, as the consistency of the colonic contents show diurnal variation, with scybala (formed pellets) forming only later during the day. Several lines of evidence, if provided like previous studies, would have supported the inferences of "toxicity" and "megacolon": (i) abdominal distention before laparotomy (ii) angry looking colonic segments (due to massive inflammation) that would have occupied the peritoneal cavity and popped out soon after laparotomy (iii) massively dilated cecum and colon (iv) retained contents in the small intestine (Figures 1, 2). The gastrointestinal transit experiments with FD70 show retention at the level of the small intestine (1). This suggests that the viral spread likely occurred to the small intestine (somewhat supported by the tissue-specific viral titers). If the myenteric ganglia damage was indeed very severe, leading to adynamism, this should have precluded contents from the cecum to enter the colon, as the entire colon would be affected and nonfunctioning. The only short terminal segment of an unfilled colon (1) suggest a mild functional constipation-like phenotype rather than "toxic megacolon." Future electrophysiologic studies and smooth muscle mechanical recordings shall help to objectively delineate and quantify the defects in intestinal motility after HSV1 genital infection.

In clinical situations, if the obstruction occurs at the level of ileum in suspected toxic megacolon, it is an ominous situation and emergency (26). For example, if surgical interventions (exploratory laparotomy) are being performed and if dilation or stasis is seen in terminal ileum, it would imply significant obstruction distally, impending colonic perforation and occurrence of subsequent peritonitis and potentially death. The fact that the mice had their obstruction reversed by administration of Miralax (polyethylene glycol) Chaudhury et al.



(1), a commonly used osmotic laxative, shows that the delayed transit in the mice after HSV infection was of a functional mild gastrointestinal motility issue of reduced transit, rather than complete obstruction as seen in toxic megacolon or adynamism encountered in intestinal pseudo-obstruction. If toxic megacolon is suspected in the clinical setting, Miralax must not be administered, as it carries the sinister risk of perforating the intestines! Toxic megacolon is a highly critical and dreaded surgical condition and the patients are very sick with high mortality rates (27). Due to irreversible colonic spastic segments, toxic megacolon may rarely be managed medically and often involve extensive surgical resections of the affected intestines (28).

The authors could not precisely delineate the aganglionic or transitional segments (1). It is possible that the ascending kind of HSV infection was stochastic, so different noncontiguous gut segments may be affected. In previous studies of models of megacolon, distinct aganglionic segments have been demonstrated (15). The present study shows representative images of myenteric ganglion in whole mounts that suggests qualitative reduction, but not complete absence (1). Hematoxylin-eosin (H&E) sections showed persistence of myenteric ganglia after HSV infection and development of colonic distention (1). The colonic walls are not thinned out, but rather comparable in thickness between wild type and infected mice. Also, the mucosa do not show any fulminant nature of colitis but intact crypt architecture and only mild mucosal infiltrate, though transmural inflammatory cell infiltrate is visualized in the colon in HSV1 infected mice which had delayed gastrointestinal transit. This type of gastrointestinal mucosal sparing with viral homing to the enteric nervous system has been reported earlier (29).

Profiling the inflammatory immune cells is the strength of this investigative group (1). These tools may be implemented in future studies examining neuroimmune interactions in the enteric nervous system. For example, an important feature in ulcerative colitis (UC) is phenotype reversal, i.e. change of the initial diarrhea-predominant symptoms to constipation (30). In fact, these UC patients are at risk of developing toxic megacolon (22). Thus, endoscopy or endoscopic insufflation is performed



with great care in these patients who present with constipation, as the procedure might perforate the colon. There are a few additional points to consider here. The authors hypothesize that neutrophils may be responsible for death of myenteric neurons (1). First, quantitative demonstration of the percent reduction of myenteric neurons has not been provided (1). Neutrophils are seen in the vicinity of the myenteric ganglia (1). However, it may be noted that mobile cells are normally present across the wall of the intestines, though their function is only scantily known (31, 32). It is obvious that some of the HSV1 infected intestinal segments had increased inflammatory cells (1). It is not clear how sick mice were identified during the experiments. For example, how were mice with potentially delayed GI transit identified and randomized to a group that received IgG or the neutrophil blocking antibody aLy6G (1). Interestingly, one of the infected mouse with prolonged luminal transit time survived 7 weeks post-infection, again indicating the mild nature of the pathophysiology involving the enteric nervous system.

The authors suggest that the HSV1 viral particles do not play a direct role in the damage of the myenteric neurons (1), but no direct evidence is provided to this critical assertion. The Circos diagram shows the expression of different viral gene transcripts (1). One of them is the  $\sim$ 140 kB pUS9, whose protein translate functions to interact with kinesin and egress viral particles out of neuronal cells (33). This may be the reason why the viral particles are seen in the extra-junctional place outside the myenteric musculomotor nerve terminal, in addition to within the neuronal cells and processes (1) (**Figure 4**). Force generating proteins and cytoskeleton are key to regulation of enteric neurotransmission (34–36). So, it is possible that HSV viral particles hijack these cell machineries like other viral particles like rabies and varicella zoster and cause myenteric neuropathy. Whether HSV particles interact with kinesin and dynein for anterograde/retrograde transport through the axons remains to be tested.

The authors report the interesting fact that the "sildenafil response proteins" are decreased in the smooth muscles and demonstrate viral particles within the smooth muscles of the muscularis externa (1). These "sildenafil response" proteins prevent degradation of the nitric oxide generated cyclic GMP (cGMP) and enhance activity of its downstream effectors, the key response elements to enteric nitrergic neuromuscular neurotransmission (37–39). It is increasingly identified that inhibitory nitrergic neurotransmission is the final common pathway mediating gastrointestinal motility (35–40). Thus, it is no surprise that the constipation like phenotype due to HSV1 infection may arise due to impairment in the myenteric nitrergic



FIGURE 3 | (I) Intraoperative view of massive dilation of colon in a subject affected with Chagas' disease Note the benign appearance of the colonic wall despite the massively dilated colon, often seen when the megacolon develops slowly over a period of time. (II) Lateral "through-view" imaging of abdomen performed after double contrast barium enema in a patient with toxic megacolon Note that the patient is positioned in knee-elbow position (inset), which is maneuvered to pass out gas. Note the enormously swollen large intestines. The particular position helps the fluid accumulate in the bottom loops of the intestines, whereas the rectum straightens out and slight stimulation, like cough etc., allows the gas to pass out and contribute to decompression of the massively enlarged colon. (III) Intra-operative view of toxic megacolon Note the lack-luster appearance of the massively dilated colon and the prominent blood vessels. The colonic swelling may impair the perfusion, and this may have additional effects on extravasation of inflammatory cells. These aspects should be factored in during examination of the inflammatory cells in the muscularis. The "toxicity" criteria in the clinical setting of megacolon is defined as per Jalan et al. (16). Additionally, functional studies like mechanical recordings and evoked potentials may help define the accurate nature of the effects of infection and inflammation on neurotransmission (17). Inflammatory bowel disease and fulminant colitis resulting from infections like *Clostridium difficile* are the common causes of this morbid condition. There are only scant reports of bowel or bladder obstruction from HSV infection (18), though with the changing epidemiology, the correlation needs to be remembered. HSV infection in particular can cause pelvic autonomic neuropathy including constipation and urinary retention as a spectrum of immune reconstitution inflammatory syndrome (IRIS) during management of HIV infection. Reproduced with permission from Munoz-Saravia (19), Panos et al.



FIGURE 4 | (I) Zoomed-out electron micrograph of a mouse with delayed gastrointestinal transit after genital HSV1 infection Note the intact nerve terminal and axon segment in the top (12 o' clock) of the panel. This varicosity shows vesicles docked at the terminal membrane (readily releasable pool, RRP), as well as recycling circulating pool (CP) and storage pool (SP). This likely indicates a normal nerve terminal. This may explain why the HSV1 infection of the enteric nervous system resulted only in a mild phenotype. Also note the HSV1 viral particles in the extracellular space exterior to the terminal. These viral particles probably transgress through the junctional space and enter the post-junctional smooth muscles and may cause defects in the post-junctional proteins like guanylyl cyclase that responds to evoked nitric oxide from the nerve terminals. (II) A cartoon depicting the possible pathways that HSV particles may affect and cause smooth muscle relaxation via upregulation of cGMP by soluble guanylyl cyclase. These pathways may be affected by viral integration in the smooth muscle genome, thus impairing smooth muscles may all be affected, resulting in impaired function of these important pelvic organs. Reproduced with permission from Khoury-Hanold et al. (1). The egress of the viral particles may be facilitated by viral proteins interacting with the neuronal cytoskeleton. NO, nitric oxide; sGC, soluble guanylyl cyclase; cGMP, cyclic guanosine monophosphate; PDE5, cGMP-specific phosphodiesterase inhibitor (sensitive to sildenafil); PKG, protein kinase G.

neurotransmission pathways by affecting both prejunctional and post-junctional mechanisms. Vesicularly released ATP mediates the gastrointestinal smooth muscle relaxation along with *de novo* synthesized nitric oxide (38). HSV has been reported to affect ATP mediated neurotransmission by upregulating CD73 (41), an ectonucleotidase that degrades extracellular junctional ATP. The transcellular spread of the viral particles from the enteric neurons to the colonic smooth muscles through the extracellular space may also explain why HSV particles may affect intraabdominal solid organs like the pancreas and cause pancreatitis (42). The transcellular travel may also facilitate spread across the pelvic space, bypassing the paravertebral or dorsal root ganglia (43).

The authors have made significant endeavor to understand the spread of the HSV1 in the pelvis and beyond (1). Because of intestinal pseudo-obstruction as well as urinary bladder outlet obstruction (1), it seems that there is retrograde transport of the virus through sensory neurons to the region of the sacral plexi, where the viral particles likely cross over and spread to multiple pelvic organs through the sacral parasympathetic outflow neurons. Understanding the mechanisms of these transneuronal pathways is an important step ahead. It remains unclear why the dorsal root ganglion remained relatively unaffected. The present study shows that the axon-pathfinding molecules are downregulated in the gastrointestinal muscularis. A component of axonal pathfinding molecule, netrin, is the main receptor for HSV1 viral entry into vaginal mucosa, and causative for encephalitis (44). The downregulation of netrin in the present study (1) could be an adaptive reaction to the ongoing infection. These discrepancies need to be sorted in future studies. It also remains unclear why only a small subset of mice develops intestinal stasis, and also how many of them developed bladder outlet obstruction. Food and water intake, as well as urine/fecal output of the mice during the advanced stages of illness has not been reported in the present study. It has earlier been reported that HSV can enter the spinal cord through all kinds of peripheral nerves and spread via trans-synaptic spread, cause an ascending disto-proximal demyelinating disease and ascend proximally to the brain, causing encephalitis (45, 46). The dynamics and extent of this spread may be multifold: the strength of the viral inoculum, the spread mechanisms, the local immune defense and the neuroviral interactions preventing lytic cycle or favoring reactivation. In the clinical setting, if there is a presentation with bladder obstruction, obstipation and signs like the vesicles of genital herpes, lumbar puncture is always indicated, as these patients are at high risk of HSV encephalitis (45, 47, 48). There has been no earlier report that these patients had mortality from toxic megacolon related to the HSV infection. Thus, epidemiologically, the gastrointestinal (GI) side effects reported in the current in vitro studies (1) may be rare in the actual clinical scenario. Urinary retention due to HSV infection is also rare, though has been reported (47, 49). The bladder outlet obstruction, due to backflow, may have impaired renal infection, caused dyselectrolytemia and death in these mice, rather than sepsis (1). No postmortem appearance of the entire gastrointestinal tract has been shown for the subset of mice who were dead after about 2 weeks of the induced HSV infection. The evidence that the mice succumbed to sepsis due to toxic colonic dilation has not been provided in the present report (1). Features such as free air or fluid in the peritoneum, or hypotension/hypothermia (**Figure 1**), would have been more supportive of evolving sepsis from severe obstruction and potential perforation of gastrointestinal luminal transit.

The topic of the current investigation is of huge epidemiologic significance, viz. the pathophysiologic consequences of HSV1 genital infection per se. The general concepts regarding HSV infection is HSV1 causes oral infection (oral herpes), whereas HSV2 causes genital herpes. The authors rightly bring out the increasing significance of the HSV1 causing genital infections (1, 50). This may be due to changing tropism of the viruses, as well as prevalent sexual practices like "putting mouth on the genital area of the sexual partner," thus causing exchange of local viruses (40, 51). Importantly, these viral infections can flare up in conditions of suppressed immunity, including pre-existing HIV (human immunodeficiency virus) or other STIs (sexually transmitted infections), pregnancy, use of corticosteroids or immune suppression due to diabetes, treatment of cancer, post-transplantation or auto-immune diseases (52-54). Key areas to explore in future shall include the extent of HSV1/2 infections on disease pathophysiology under controlled conditions of immune compromise (54, 55). Studies have shown that enterocolitis that develops in chronic megacolon like Hirschsprung's disease importantly involve changes in luminal microbiome and mycobiome (56-58). An important area to consider is whether HSV infection may alter luminal microbes and fungal composition, and whether that may be causative of enteric neuronal dysfunction.

All classes of herpesviridae have been reported to infect myenteric neurons, including varicella zoster, cytomegalovirus and herpesvirus, including HSV1 and HSV2 (59-62). In such respect, the present study by Khoury-Hanold et al. (1) do not really usher a novel aspect. There have been some recent reports of viral diagnostic methods which can identify these organisms (63-65). However, the entire field of direct viral detection is in its infancy and these are not routinely employed in the care setting. There are reports of viruses that persist in the myenteric neurons and cause major gastrointestinal dysmotility, for example bornavirus and achalasia (66). These associations have also been reported with HSV (67). However, very little is known regarding the mechanisms of the latency of the viruses within the neurons, their genomic integration or their clearance mechanisms by the immune system and ultimate effect on enteric neurotransmission (68). In immunocompromised states, the "viruses may show their true color."

The gastrointestinal tract has a massive immune surveillance system due to the challenges of microbes consumed through ingested food. Increasingly, with sexual preferences like sodomy practiced during MSM (men sex with men), the outlet of the gastrointestinal tract is faced with an ever-increasing challenge of tackling microrganisms including viruses (40, 69). Most often, the distal immune system will tackle these infectious agents. But viruses may "walk back" through neurons and thereafter spread

far and wide, wreaking havoc. When tell-tales of herpes infection like vesicles are seen, it helps in instituting antiviral prophylaxis and treatment. However, atypical and subclinical presentations can occur, and herpes may manifest without vesicles (46, 70). The present study demonstrates that both HSV 1 and 2 cause neuropathy in pelvic organs (1). Future studies can systematically examine the effect of viruses on nerve densities in the wall of the intestine and any frank morphologic abnormality, or whether the disease is "functional" in manifestation. Both these strains of viruses are neurotropic and can track back to the central nervous system via multiple routes (1, 46), causing life-threatening infections. The autonomic neuropathy can cause multisystem involvement and significant impairment of organ function like delayed gastrointestinal transit and impaired outflow of urine. In our opinion, this significant report of their elaborate investigations by Khoury-Hanold et al.(1) brings forth this hugely

## REFERENCES

- Khoury-Hanold W, Yordy B, Kong P, Kong Y, Ge W, Szigeti-Buck K, et al. Viral spread to enteric neurons links genital HSV-1 infection to toxic megacolon and lethality. *Cell Host Microbe* (2016) 19:788–99. doi: 10.1016/j.chom.2016.05.008
- Gariepy CE, Cass DT, Yanagisawa M. Null mutation of endothelin receptor type B gene in spotting lethal rats causes aganglionic megacolon and white coat color. *Proc Natl Acad Sci USA*. (1996) 93:867–72. doi: 10.1073/pnas.93.2.867
- Lipman NS, Wardrip CL, Yuan CS, Coventry S, Bunte RM, Li X. Familial megacecum and colon in the rat: a new model of gastrointestinal neuromuscular dysfunction. *Lab Anim Sci.* (1998) 48:243–52.
- 4. Zimmer J, Puri P. Knockout mouse models of Hirschsprung's disease. *Pediatr* Surg Int. (2015) 31:787–94. doi: 10.1007/s00383-015-3747-3
- Collins SM. The immunomodulation of enteric neuromuscular function: implications for motility and inflammatory disorders. *Gastroenterology* (1996) 111:1683–99.
- Koenigsknecht MJ, Theriot CM, Bergin IL, Schumacher CA, Schloss PD, Young VB. Dynamics and establishment of *Clostridium difficile* infection in the murine gastrointestinal tract. *Infect Immun.* (2015) 83:934–41. doi: 10.1128/IAI.02768-14
- Schmidt DJ, Beamer G, Tremblay JM, Steele JA, Kim HB, Wang Y, et al. A tetraspecific VHH-based neutralizing antibody modifies disease outcome in three animal models of *Clostridium difficile*. *Infect Clin Vaccine Immunol*. (2016). 23:774–84. doi: 10.1128/CVI.00730-15
- Kang SS, Bloom SM, Norian LA, Geske MJ, Flavell RA, Stappenbeck TS, et al. An antibiotic-responsive mouse model of fulminant ulcerative colitis. *PLoS Med.* (2008) 5:e41. doi: 10.1371/journal.pmed.0050041
- Vallance BA, Gunawan MI, Hewlett B, Bercik P, Van Kampen C, Galeazzi F, et al. TGF-beta1 gene transfer to the mouse colon leads to intestinal fibrosis. *Am J Physiol Gastrointest Liver Physiol.* (2005) 289:G116–28. doi: 10.1152/ajpgi.00051.2005
- Friebe A, Mergia E, Dangel O, Lange A, Koesling D. Fatal gastrointestinal obstruction and hypertension in mice lacking nitric oxide-sensitive guanylyl cyclase. *Proc Natl Acad Sci USA*. (2007) 104:7699–704. doi: 10.1073/pnas.0609778104
- Khalil HA, Kobayashi M, Rana P, Wagner JP, Scott A, Yoo J, Dunn JC. Mouse model of endoscopically ablated enteric nervous system. J. Surg. Res. (2016) 200:117–21. doi: 10.1016/j.jss.2015.07.034
- 12. Jelicks LA. Imaging the gastrointestinal tract of small animals. J Neuroparasitol. (2010). 1:N100504.
- Kim H, Kang K, Ekram MB, Roh TY, Kim J. Aebp2 as an epigenetic regulator for neural crest cells. *PLoS ONE* (2011) 6:e25. doi: 10.1371/journal.pone.0025174
- 14. McCallion AS, Stames E, Conlon RA, Chakravarti, A. Phenotype variation in two-locus mouse models of Hirschsprung disease: tissue-specific interaction

important epidemiologic issue into the limelight of systematic scientific investigation.

## **AUTHOR CONTRIBUTIONS**

AC: conceptualized and drafted manuscript; literature review; VD, MC, AJ, MK, KP, GJ, AR, BR, SRK, SF, SK, NSS, SM: planning of manuscript and clinical and technical collaborative discussions. All authors approved final version of manuscript.

# ACKNOWLEDGMENTS

The authors wish to thank Dr. Akiko Iwasaki for prompt sharing of the article. The authors thank Sushmita Biswas for helpful discussion on designing the cartoon.

between Ret and Ednrb. Proc Natl Acad Sci USA. (2003) 100:1826-31. doi: 10.1073/pnas.0337540100

- Obermayr F, Hotta R, Enomoto H, Young, HM. Development and developmental disorders of the enteric nervous system. *Nat Rev Gastroenterol Hepatol.* (2012) 10:43–57. doi: 10.1038/nrgastro.2012.234
- Jalan KN, Sircus W, Card WI, Falconer CW, Bruce CB, Crean GP, et al. An experience of ulcerative colitis. I. Toxic dilation in 55 cases. *Gastroenterology* (1969) 57:68–82.
- Mourelle M, Vilaseca J, Guarner F, Salas A, Malagelada JR Toxic dilatation of colon in a rat model of colitis is linked to an inducible form of nitric oxide synthase. *Am J Physiol.* (1996) 270(3 pt 1):G425–30. doi: 10.1152/ajpgi.1996.270.3.G425
- Eberhardt O, Küker W, Dichgans, J, Weller, M. HSV-2 sacral radiculitis (Elsberg syndrome). *Neurology* (2004) 63:758–9. doi: 10.1212/01.WNL.0000134652.51657.10
- Munoz-Saravia SG, Haberland A, Wallukat G, Schimke I. Chronic Chagas' heart disease – from pathogenesis to treatment regimes. *Appl Cardiopulmonary Pathophysiol.* (2012). 16:55–81.
- 20. Panos MZ, Wood MJ, Asquith, P. Toxic megacolon: the knee-elbow position relieves bowel distension. *Gut* (1993) 34:1726–7. doi: 10.1136/gut.34.12.1726
- Alterman D, Grandas O, Goldman M, Solla J. Surgical management of *Clostridium difficile* associated colitis with toxic megacolon in a transplant patient: Case Report and Review of Literature. *Internet J Surg.* (2009) 22. Available online at: https://print.ispub.com/api/0/ispub-article/13061
- Autenrieth DM, Baumgart, DC. Toxic megacolon. Inflamm Bowel Dis. (2012) 18:584–91. doi: 10.1002/ibd.21847
- Choi JS, Lim JS, Kim H, Choi JY, Kim MJ, Kim NK, et al. Colonic pseudoobstruction: CT findings. Am J Roentgenol. (2008) 190:1521–6. doi: 10.2214/AJR.07.3159
- 24. Sheth SG, LaMont, JT. Toxic megacolon. Lancet. (1998) 351:509–13. doi: 10.1016/S0140-6736(97)10475-5
- O'Dwyer RH, Acosta A, Camilleri M, Burton D, Busciglio I, Bharucha AE. Clinical features and colonic motor disturbances in chronic megacolon in adults. *Dig Dis Sci.* (2015) 60:2398–407. doi: 10.1007/s10620-015-3645-5
- Hokama A, Ohira T, Kishimoto K, Kinjo F, Fujita J. Impending megacolon: small bowel distension as a predictor of toxic megacolon in ulcerative colitis. *Intern Emerg Med.* (2012) 7:487–8. doi: 10.1007/s11739-012-0834-5
- Martin JR, Stoner, GL. The nervous system in genital herpes simplex virus type 2 infections in mice. Lethal panmyelitis or nonlethal demyelinative myelitis or meningitis. *Lab Invest.* (1984) 51:556–66.
- Strong SA. Management of acute colitis and toxic megacolon. Clin Colon Rectal Surg. (2010) 23:274–84. doi: 10.1055/s-0030-1268254
- Gesser RM, Valyi-Nagy T, Fraser NW, Altschuler SM. Oral inoculation of SCID mice with an attenuated herpes simplex virus-1 strain causes persistent enteric nervous system infection and gastric ulcers without direct mucosal infection. *Lab Invest.* (1995) 73:880–9.

- Rao SS, Holdsworth CD, Read, NW. Symptoms and stool patterns in patients with ulcerative colitis. *Gut* (1988) 29:342–5. doi: 10.1136/gut.29.3.342
- Kin NW, Sanders, VM. It takes nerve to tell T and B cells what to do. *J Leukoc Biol.* (2006) 79:1093–104. doi: 10.1189/jlb.1105625
- 32. Phillips RJ, Powley TL. Macrophages associated with the intrinsic and extrinsic autonomic innervation of the rat gastrointestinal tract. *Auton Neurosci.* (2012) 169:12–27. doi: 10.1016/j.autneu.2012.02.004
- Diefenbach RJ, Davis A, Miranda-Saksena M, Fernandez MA, Kelly BJ, Jones CA, et al. The basic domain of herpes simplex virus 1 pUS9 recruits kinesin-1 to facilitate egress from neurons. J Virol. (2015) 90:2102–11. doi: 10.1128/JVI.03041-15
- Chaudhury A. Molecular handoffs in nitrergic neurotransmission. Front Med. (2014) 1:8. doi: 10.3389/fmed.2014.00008
- Chaudhury A. Evidence for dual pathway for nitrergic neuromuscular transmission in doubt: evidence favors lack of role of ICC. *Gastroenterology* (2013) 145:1160–1. doi: 10.1053/j.gastro.2013.09.039
- Chaudhury A. Furthering the debate on the role of interstitial cells of Cajal (ICCs) in enteric inhibitory neuromuscular neurotransmission. *Am J Physiol Cell Physiol.* (2016). 311:C479–81. doi: 10.1152/ajpcell.00067.2016
- 37. Chaudhury A. Response: "Commentary: a hypothesis for examining skeletal muscle biopsy-derived sarcolemmal nNOS $\mu$  as surrogate for enteric nNOS $\alpha$  function". nNOS(skeletal muscle) may be evidentiary for enteric NO-transmission despite nNOS $\mu/\alpha$  differences. *Front Med.* (2016) 3:4. doi: 10.3389/fmed.2016.00004
- Chaudhury, A. A hypothesis for examining skeletal muscle biopsy-derived sarcolemmal nNOSµ as surrogate for enteric nNOSα function. *Front Med.* (2015) 2:48. doi: 10.3389/fmed.2015.00048
- Chaudhury A. No role of interstitial cells of Cajal (ICCs) in genesis of inhibitory junction potentials (IJPs): evidence from a novel mouse model of genomic ICC depletion. *Nature* (2014) 464:466. doi: 10.1038/464466a
- Chaudhury A. VIP in HIV Diarrhea: finding links for the "Slim Disease". Front Physiol. (2015) 6:402. doi: 10.3389/fphys.2015.00402
- Zoppellaro C, Bin A, Brun P, Banzato S, Macchi V, Castagliuolo I, et al. Adenosine-mediated enteric neuromuscular function is affected during herpes simplex virus type 1 infection of rat enteric nervous system. *PLoS ONE* (2013) 8:e72648. doi: 10.1371/journal.pone.0072648
- Chen YG, Huang KL, Peng, CK. A man with diffuse vesicular rash and epigastric pain. *Gastroenterology*. (2013) 145:e9–10. doi: 10.1053/j.gastro.2013.06.039
- Ohashi M, Bertke AS, Patel A, Krause, PR. Spread of herpes simplex virus to the spinal cord is independent of spread to dorsal root ganglia. *J Virol.* (2011) 85:3030–2. doi: 10.1128/JVI.02426-10
- Kopp SJ, Banisadr G, Glajch K, Maurer UE, Grünewald K, Miller RJ, et al. Infection of neurons and encephalitis after intracranial inoculation of herpes simplex virus requires the entry receptor nectin-1. *Proc Natl Acad Sci USA*. (2009) 106:17916–20.
- Martin JR, Suzuki, S. Inflammatory sensory polyradiculopathy and reactivated peripheral nervous system infection in a genital herpes model. *J Neurol Sci.* (1987) 79:155–71. doi: 10.1016/0022-510X(87)90270-X
- 46. Ugolini G. Transneuronal transfer of herpes simplex virus type 1 (HSV 1) from mixed limb nerves to the CNS. I. Sequence of transfer from sensory, motor, and sympathetic nerve fibres to the spinal cord. *J Comp Neurol.* (1992) 326:527–48. doi: 10.1002/cne.903260404
- Berger JR, Houff, S. Neurological complications of herpes simplex virus type 2 infection. Arch Neurol. (2008) 65:596–600. doi: 10.1001/archneur.65.5.596
- Goodell SE, Quinn TC, Mkrtichian E, Schuffler MD, Holmes KK, Corey L. Herpes simplex virus proctitis in homosexual men. Clinical, sigmoidoscopic, and histopathological features. *N Engl J Med.* (1983) 308:868– 71. doi: 10.1056/NEJM198304143081503
- Caplan LR, Kleeman FJ, Berg, S. Urinary retention probably secondary to herpes genitalis. N Engl J Med. (1977) 297:920–1. doi: 10.1056/NEJM197710272971708
- 50. Gupta R, Warren T, Wald, A. Genital herpes. Lancet (2007) 370:2127-37.
- Cherpes TL, Meyn LA, Hillier SL. Cunnilingus and vaginal intercourse are risk factors for herpes simplex virus type 1 acquisition in women. *Sex Transm Dis.* (2005) 32:84–9. doi: 10.1097/01.olq.0000151414.64297.46
- Brown ZA, Selke S, Zeh J, Kopelman J, Maslow A, Ashley RL, et al. The acquisition of herpes simplex virus during pregnancy. N Engl J Med. (1997) 337:509–15. doi: 10.1056/NEJM199708213370801

- Corey, L. Synergistic copathogens–HIV-1 and HSV-2. N Engl J Med. 356:854–6. Erratum in (2007) N Engl J Med. (2007) 356:1487. doi: 10.1056/NEJMe068302
- Takase H, Yamamura E, Murakami Y, Ikeuchi T, Osada Y. Gastrointestinal invasion by herpes simplex virus type 1 inoculated cutaneously into the immunosuppressed mice. *Arch Virol.* (1994) 134:97–107. doi: 10.1007/BF01379110
- Sanjuan NA, Lascano EF. Autonomic nervous system involvement in experimental genital infection by herpes simplex virus type 2. Arch Virol. (1986) 91:329–39. doi: 10.1007/BF01314291
- Gosain A, Brinkman AS. Hirschsprung's associated enterocolitis. Curr Opin Pediatr. (2015) 27:364–9. doi: 10.1097/MOP.00000000000210
- 57. Frykman PK, Nordenskjöld A, Kawaguchi A, Hui TT, Granström AL, Cheng Z, et al. Characterization of bacterial and fungal microbiome in children with hirschsprung disease with and without a history of enterocolitis: a multicenter study. *PLoS ONE* (2015) 10:e0124172. doi: 10.1371/journal.pone.0124172
- Iliev ID, Funari VA, Taylor KD, Nguyen Q, Reyes CN, Strom SP, et al. Interactions between commensal fungi and the C-type lectin receptor Dectin-1 influence colitis. *Science* (2012) 336:1314–7. doi: 10.1126/science.1221789
- Brun P, Giron MC, Zoppellaro C, Bin A, Porzionato A, De Caro R, et al. Herpes simplex virus type 1 infection of the rat enteric nervous system evokes small-bowel neuromuscular abnormalities. *Gastroenterology* (2010) 138:1790–801. doi: 10.1053/j.gastro.2010.01.036
- Chen JJ, Gershon AA, Li Z, Cowles RA, Gershon MD. Varicella zoster virus (VZV) infects and establishes latency in enteric neurons. *J Neurovirol*. (2011) 17:578–89. doi: 10.1007/s13365-011-0070-1
- Debinski HS, Kamm MA, Talbot IC, Khan G, Kangro HO, Jeffries DJ. DNA viruses in the pathogenesis of sporadic chronic idiopathic intestinal pseudoobstruction. *Gut* (1997) 41:100–6. doi: 10.1136/gut.41.1.100
- 62. Gesser RM, Koo SC. Oral inoculation with herpes simplex virus type 1 infects enteric neuron and mucosal nerve fibers within the gastrointestinal tract in mice. *J Virol*. (1996) 70:4097–102.
- Gershon AA, Chen J, Gershon MD. Use of saliva to identify varicella zoster virus infection of the Gut. *Clin Infect Dis.* (2015) 61:536–44. doi: 10.1093/cid/civ320
- Lau KW, McCaughey C, Coyle PV, Murray LJ, Johnston BT. Enhanced reactivity of peripheral blood immune cells to HSV-1 in primary achalasia. *Scand J Gastroenterol.* (2010) 45:806–13. doi: 10.3109/00365521003587804
- Xu GJ, Kula T, Xu Q, Li MZ, Vernon SD, Ndung'u T, et al. Viral immunology. comprehensive serological profiling of human populations using a synthetic human virome. *Science* (2015). 348:aaa0698. doi: 10.1126/science.aaa0698
- Goyal RK, Chaudhury, A. Pathogenesis of achalasia: lessons from mutant mice. *Gastroenterology* (2010) 139:1086–90. doi: 10.1053/j.gastro.2010. 08.013
- Boeckxstaens, GE. Achalasia: virus-induced euthanasia of neurons? *Am J Gastroenterol.* (2008) 103:1610–2. doi: 10.1111/j.1572-0241.2008. 01967.x
- Knipe DM, Raja P, Lee, JS. Clues to mechanisms of herpesviral latent infection and potential cures. *Proc Natl Acad Sci USA*. (2015) 112:11993–4. doi: 10.1073/pnas.1516224112
- Quinn TC, Stamm WE, Goodell SE, Mkrtichian E, Benedetti J, Corey L, et al. The polymicrobial origin of intestinal infections in homosexual men. N Engl J Med. (1983) 309:576–82. doi: 10.1056/NEJM198309083091002
- Wald A, Zeh J, Selke S, Ashley RL, Corey, L. Virologic characteristics of subclinical and symptomatic genital herpes infections. *N Engl J Med.* (1995) 333:770–5.

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Chaudhury, Dendi, Chaudhury, Jain, Kasarla, Panuganti, Jain, Ramanujam, Rena, Koyagura, Fogla, Kumar, Shekhawat and Maddur. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.